

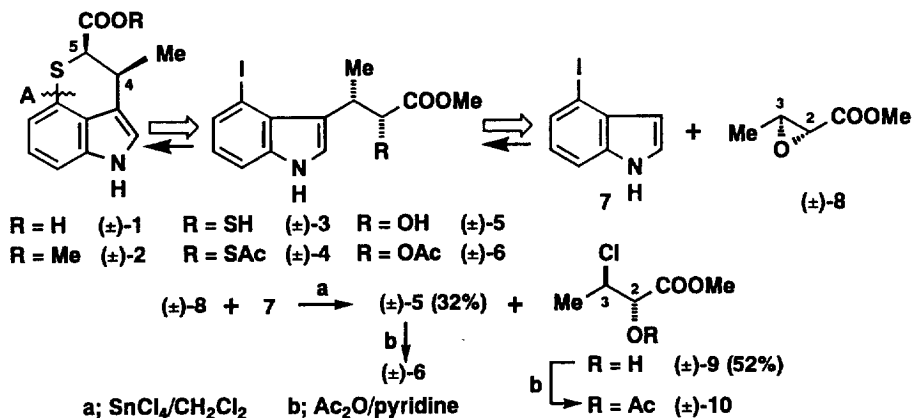
New total synthesis of (–)- and (+)-chuangxinmycins

Keisuke Kato, Machiko Ono and Hiroyuki Akita*

School of Pharmaceutical Science, Toho University, 2–2–1, Miyama, Funabashi, Chiba 274, Japan

Abstract: Stereoselective total syntheses of (–)-(4*S*,5*R*)- and (+)-(4*R*,5*S*)-chuangxinmycins **1** were achieved based on the enzymatic syntheses of (2*R*,3*S*)- and (2*S*,3*R*)-epoxy butanoates **8**, respectively. Chiral intermediates such as (2*R*,3*S*)- and (2*S*,3*R*)-2-hydroxy-3-(4'-iodoindol-3'-yl)butanoate **5** for the chiral synthesis of (–)- and (+)-**1** were also obtained by the enantioselective hydrolysis of the corresponding acetate **6** by lipase. © 1997 Elsevier Science Ltd

Chuangxinmycin **1**, isolated from *Actinoplanes tsinanensis* n. sp. in China, exhibits *in vitro* against a number of Gram-positive and Gram-negative bacteria. This material was reported to be active in mice against *Escherichia coli* and *Shigella dysenteria* infections *in vivo*, and effective in the treatment of septicemia, urinary, and biliary infections caused by *E. coli* in preliminary clinical results.¹



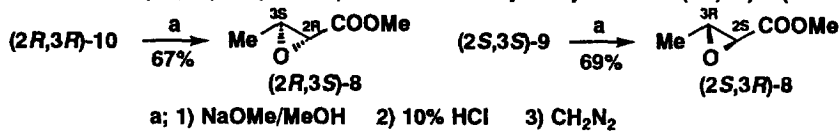
The relative structure of **1** was confirmed by racemic synthesis² and the absolute configurations were determined as 4*S*,5*R* based on the degradation study of the natural product **1**³ and resolution of (±)-**1** with *S*-(-)- α -methylbenzylamine.⁴ In preceding paper, we reported a highly stereoselective synthesis of (±)-**1** *via* pathway A directed toward chiral synthesis involving two characteristic approaches to this problem.⁵ The first one is the stereoselective conversion of (±)-(2,3)-*syn*-2-hydroxy-3-(4'-iodoindol-3'-yl)butanoate **5** into the (±)-(2,3)-*syn*-2-mercapto ester **3** *via* the corresponding 2-thioacetoxy ester **4** with retention of C₂-stereochemistry in (±)-**5**. The second one is the palladium-catalyzed cyclization of indolyl iodide and the internal C₂-thiol group of the substrate (±)-**3** to give the (±)-methyl ester **2** of (±)-chuangxinmycin **1** without isomerization at the C₅-position in high yield. The synthesis of (±)-**5** was achieved by the reaction of 4'-iodoindole **7**⁵ and (±)-*trans*-(2,3)-epoxy butanoate **8** in the presence of SnCl₄ along with nucleophilic displacement with inversion at the C-3 carbon of the coordinated epoxide. We now report a highly stereoselective synthesis of both enantiomers of chuangxinmycin **1** by the following two approaches as a key step. One is the synthesis of enantiomerically active (2*R*,3*S*)- or (2*S*,3*R*)-**8** based on the asymmetric hydrolysis of (±)-(2,3)-*anti*-acetoxy-3-chloro butanoate **10** using lipase (Table 1). The other is the syntheses of enantiomerically active (2*R*,3*S*)- or (2*S*,3*R*)-**3** and

* Corresponding author. Email: akita@phar.toho-u.ac.jp

Table 1.

Entry	Substrate (g, %ee)	Time(days)	Products	
			% (%ee)	% (%ee)
1	(±)-10 (14)	5	(2R,3R)-10; 45 (87)	(2S,3S)-9; 40 (89)
2	(2R,3R)-10 (6.3, 87)	3	(2R,3R)-10; 84 (>99)	(2S,3S)-9; 12 (0)
3	(2S,3S)-10 (4.4, 89)*	3	(2S,3S)-10; 12 (27)	(2S,3S)-9; 68 (>99)

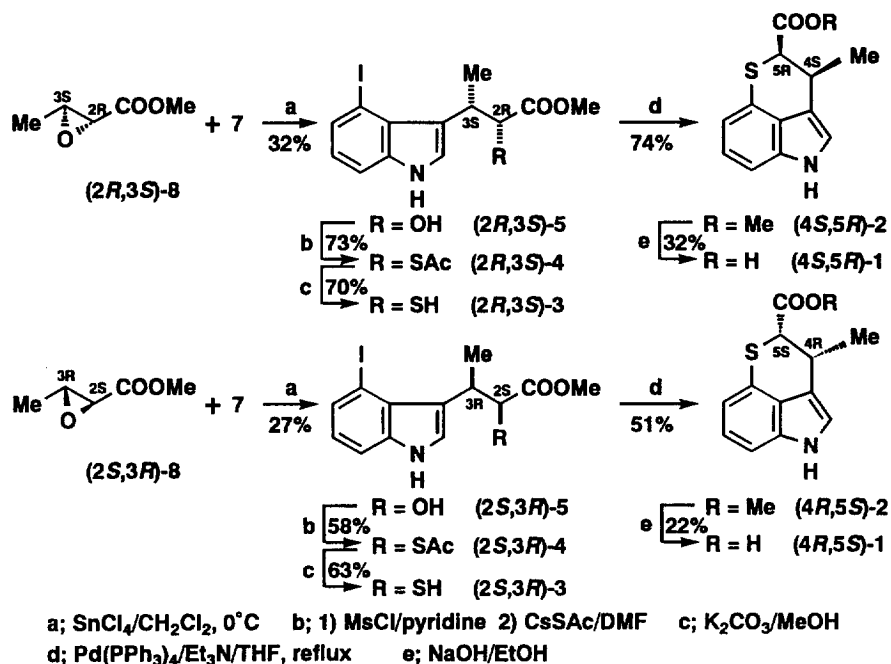
* The substrate (2S,3S)-10 (89% ee) was obtained by acetylation of (2S,3S)-9 (89% ee).



(2R,3S)- or (2S,3R)-5 based on the asymmetric hydrolysis of the corresponding acetates (±)-4 or (±)-6 using lipase, respectively. The substrates (±)-6 and (±)-10 for enzymatic reaction were obtained by acetylation of the reported 2-hydroxy esters (±)-5 and (±)-9, respectively, which were provided by the reaction of 7 and (±)-8 in the presence of SnCl₄.⁵

Initially, (±)-10 was subjected to screening experiments using several kinds of commercially available lipases in isopropyl ether saturated with water. Among them, lipase "Amano P" from *Pseudomonas* sp. was found to give the (2S,3S)-2-hydroxy ester 9⁶ (40%, 89% ee) and the unchanged (2R,3R)-10 (45%, 87% ee)(entry 1). The (2R,3R)-10 having 87% ee was again subjected to the enzymatic hydrolysis to afford the enantiomerically pure (2R,3R)-10 ([α]_D +7.6 (c=1.50, CHCl₃)) in 84% yield (entry 2), which was consistent with the reported (2R,3R)-10⁷ ([α]_D +8.3 (c=3.0, CHCl₃): corresponds to >99% ee). On the contrary, the 89% enantiomeric excess of (2S,3S)-10 was subjected to the enzymatic hydrolysis to provide the enantiomerically pure (2S,3S)-9 ([α]_D +33.2 (c=0.85, CHCl₃)) in 68% yield (entry 3). The enantiomeric excess of (+)-10 and (+)-9 were calculated based on NMR (400 MHz) data of the corresponding (R)-(+)-α-methoxy-α-trifluoromethyl-phenylacetates⁸ ((R)-MTPA esters: (R)-MTPA ester from (+)-10, δ 3.836, COOMe; (R)-MTPA ester from (+)-9, δ 3.785, COOMe). Thus obtained (2R,3R)-10 was treated with NaOMe followed by the subsequent acid treatment and esterification with CH₂N₂ to provide the enantiomerically pure (2R,3S)-epoxy butanoate (8) in 67% overall yield. Likewise, the enantiomerically pure (2S,3S)-9 was also converted to the (2S,3R)-8 in 69% overall yield. The reaction of 7⁵ and (2R,3S)-8 in the presence of SnCl₄ afforded (2R,3S)-5 (32% yield, mp 52°C, [α]_D +7.89 (c=0.5, CHCl₃)), which was treated with MsCl in pyridine followed by treatment with CsSAc to provide (2R,3S)-2-thioacetoxy ester 4([α]_D +84.9 (c=0.94, CHCl₃)) in 73% overall yield with complete retention of C₂-stereochemistry. Deacetylation of (2R,3S)-4 with K₂CO₃ in MeOH provided (2R,3S)-2-mercapto ester 3 (70% yield, [α]_D +23.3 (c=0.3, CHCl₃)), which was subjected to the Pd(PPh₃)₄ mediated cyclization in the presence of Et₃N to give the (4S,5R)-cis-methyl ester 2 (mp 130°C, [α]_D -124.0 (c=0.45, CHCl₃)) in 74% yield. An alkaline hydrolysis of (4S,5R)-2 was carried out by the reported procedure^{2c} to provide the natural chuangxinmycin (4S,5R)-1 (mp 192–193°C, [α]_D -26.0 (c=0.2, 95% EtOH)), which is consistent with the reported (4S,5R)-1 (mp 192–192.5°C,^{2a} mp 184–187°C,⁴ [α]_D -29.0 (95% EtOH)⁴). Likewise, the enantiomerically pure (2S,3R)-8 was converted to the (4R,5S)-1 (mp 181–182°C, [α]_D +27.4 (c=0.42, 95% EtOH)) via (2S,3R)-5 (mp 55°C, [α]_D -7.78 (c=0.9, CHCl₃)), (2S,3R)-4 ([α]_D -87.8 (c=0.79, CHCl₃)), (2S,3R)-3 ([α]_D -23.1 (c=0.77, CHCl₃)) and (4R,5S)-2 (mp 135°C, [α]_D +124.1 (c=0.59,

CHCl₃). The physical data of the present (4*R*,5*S*)-1 was identical with those (mp 181–184°C, [α]_D²⁰ +29 (95% EtOH)) of the reported (4*R*,5*S*)-1.⁴



Then the enantioselective hydrolysis of (±)-2-thioacetoxyster 4⁵ and (±)-2-acetoxyster 6 using lipase “OF-360” from *Candida rugosa* in the mixed solvent (cyclohexane: *i*-Pr₂O=19:1) saturated with water was carried out and the results are shown in Table 2. Enantiomeric excess (ee) of the products was estimated by HPLC on a CHIRALCEL OD (250×4.6 mm) column and absolute structure of the products was confirmed by a direct comparison with the retention times of the above-mentioned standard samples by HPLC analysis. Enzymatic hydrolysis of (±)-4 gave the compound (2*S*,3*R*)-3 (15%, 68% ee) and the unchanged acetate (2*R*,3*S*)-4 (74%, 14% ee) (entry 1), while enzymatic hydrolysis of (±)-6 afforded the 91% ee of (2*S*,3*R*)-5 (35%) and the unchanged acetate (2*R*,3*S*)-6 (64%, 51% ee) (entry 2). The recovered (2*R*,3*S*)-6 having 51% ee and 87% ee was again subjected to the enzymatic reaction to provide 87% ee and 97% ee of (2*R*,3*S*)-6, respectively (entries 3 and 4). Enrichment of ee of (2*S*,3*R*)-5 (>99% ee) was also achieved by the repeated enzymatic reaction of (2*S*,3*R*)-6 (91% ee) (entry 5).

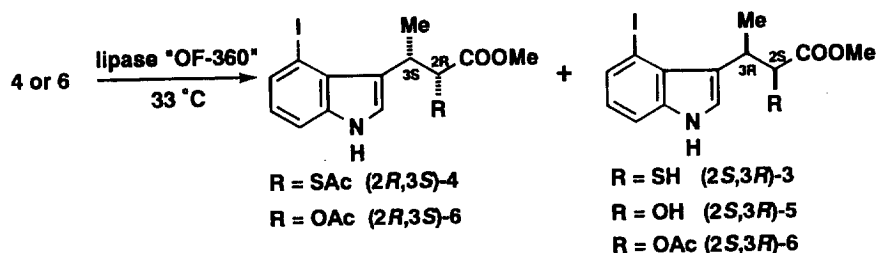
Acknowledgements

The authors are grateful to Amano Pharmaceutical Co., Ltd, (Japan) and Meito Sangyo Co., Ltd, (Japan) for providing the lipases “Amano P” and “OF-360”, respectively. This work was supported by a grant for the “Biodesign Research Program” from The Institute of Physical and Chemical Research (RIKEN, Japan) to H. A.

References

- Institute of Materia Medica, Chinese Academy of Medical Sciences, *Scientia Sin.*, **1977**, 20, 106–112.
- a) Chang Ghi-ping, Hsu Hsien-Dong, Huang Lung-Chen, Lin Yin-Ching, Li Ho-Shui, Yu Chi-Li, Chao Chun-Lan, *Acta Chim. Sin.*, **1976**, 34, 133–142. b) Kozikowski A. P., Greco M. N., *J. Am. Chem. Soc.*, **1980**, 102, 1165–1166. c) Kozikowski A. P., Greco M. N., Springer J. P., *J. Am. Chem. Soc.*, **1982**, 104, 7622–7626. d) Matsumoto M., Watanabe N., *Heterocycles*, **1987**, 26, 1775–1778. e) Murase M., Koike T., Moriya T., Tobinaga S., *Chem. Pharm. Bull.*, **1987**, 35, 2656–2660. f)

Table 2.



Entry	Substrate (mg, %ee)	Time(days)	Products	
			% (%ee)	% (%ee)
1	(±)-4 (97)	3	(2R,3S)-4; 74 (14) ¹⁾	(2S,3R)-3; 15 (68) ¹⁾
2	(±)-6 (700)	4	(2R,3S)-6; 64 (51)	(2S,3R)-5; 35 (91)
3	(2S,3S)-6 (440, 51)	3.5	(2R,3S)-6; 75 (87)	(2S,3R)-5; 23 (67)
4	(2R,3S)-6 (330, 87)	3	(2R,3S)-6; 91 (97)	(2S,3R)-5; 5 (12)
5	(2S,3R)-6 (240, 91) ²⁾	6.5	(2S,3R)-6; 24 (63)	(2S,3R)-5; 62 (>99)

1) The ee was calculated based on NMR (400 MHz) data of the corresponding (R)-(+)-MTPA ester.

2) The substrate (2S,3R)-6 (91% ee) was obtained by acetylation of (2S,3R)-5 (91% ee).

Michael J. D., Timothy J. M., David A. W., Alexandra M. Z. Slawin, David J. W., *J. Chem. Soc. Perkin I*, **1992**, 323–325. g) Ishibashi H., Tabata T., Hanaoka K., Iriyama H., Akamatsu S., Ikeda M., *Tetrahedron Lett.*, **1993**, 34, 489–492. h) Ishibashi H., Akamatsu S., Iriyama H., Ikeda M., *Chem. Pharm. Bull.*, **1994**, 42, 271–276.

- Gu Zhi-Ping, Liang Xiao-Tian, *Acta Chim. Sin.*, **1985**, 43, 250–256.
- Guo Xia-Ling, Zhang Zhi-Ping, *Acta Pharm. Sin.*, **1987**, 22, 671–678.
- Kato K., Ono M., Akita H., *Tetrahedron Lett.*, **1997**, 38, 1805–1808.
- Satisfactory analytical data were obtained for all new compounds.
- Akita H., Kawaguchi T., Enoki Y., Oishi T., *Chem. Pharm. Bull.*, **1990**, 38, 323–328.
- a) Dale J. A., Dull D. L., Mosher H. S., *J. Org. Chem.*, **1969**, 34, 2543–2549. b) Dale J. A., Mosher H. S., *J. Am. Chem. Soc.*, **1973**, 95, 512–519.

(Received in Japan 21 May 1997)